

Development of modified QuEChERS extraction methods for the analysis of PAHs in high fat content fish and smoked salmon

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Rationale:

Polycyclic aromatic hydrocarbons (PAHs) are a class of pollutants that can contaminate biotic tissue exposed to pyro- and/or petrogenic processes (i.e biomass combustion and oil spills). They are composed principally of carbon and hydrogen atoms arranged into greater than two aromatic rings and have $\log K_{ow}$ s ranging from 3 – 7. It is known that some PAHs, such as benzo[a]pyrene, possess genotoxic, mutagenic and carcinogenic properties. As such, user-friendly analytical methods that allow for simple, efficient and robust quantification of PAHs are of great interest.

Chemicals of Concern

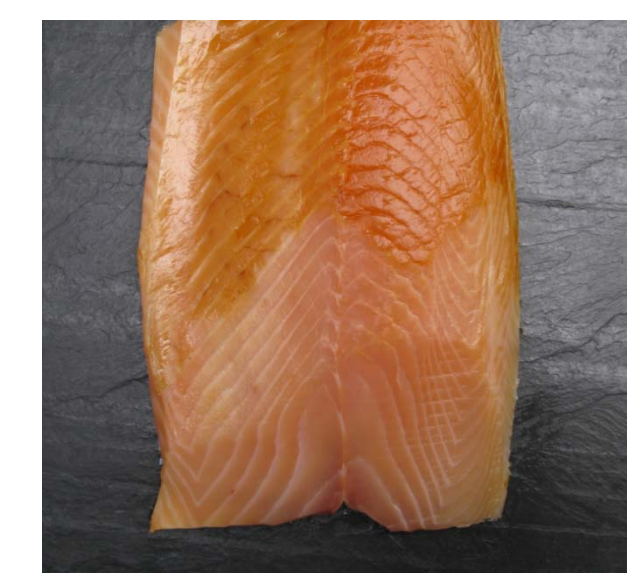
Analytical Methods

Sample preparation using QuEChERS (Quick, Easy, Cheap, Effective, Rugged and Safe) approaches are becoming increasingly popular for chemical extraction and clean-up from fruits and vegetables. Originally developed for pesticides, QuEChERS methods traditionally involve sample extraction into acetonitrile, subsequent liquid-liquid partitioning following addition of $MgSO_4$ and $NaCl$ and dispersive SPE clean-up^a. However, little information has been published on the applicability of these techniques for analyzing highly hydrophobic contaminants in high fat content bio-matrices such as smoked meats or oiled organisms.

OBJECTIVE: Develop and validate a sensitive QuEChERS sample preparation method for identifying and quantifying PAHs in high fat bio-matrices

QuEChERS Sample Preparation Methodology

Samples: Commercially available smoked salmon with 3, 8 and 11 % fat, as reported in product nutrition fact labels, were homogenized via freeze fracture with liquid N_2 in a food processor.



Extraction: Solid-liquid extraction into an organic solvent, liquid-liquid partitioning with QuEChERS EN or AOAC extraction salts and centrifugation

Clean-up: QuEChERS fatty sample d-SPE tubes and centrifugation

Chemical analysis: GC-MS equipped with a DB-5MS column, inlet temp: 300° C, flow rate: 1 mL/min, ionization: electron impact (70 eV), SIM mode, total run time - 36 min.

Quantitative PAHs

Compound	DBS R _t (min)	Target compound SIM ions (m/z)		R ²	MDL (µg kg ⁻¹)	FDA levels of concern (µg kg ⁻¹)
		Quant	Confirm			
Perylene D-12	25.72	264	260, 265	Internal standard	-	-
Naphthalene	8.77	128	127, 129	0.999	2	32,700
2-Methylnaphthalene	10.30	142	141, 115	0.999	2	-
1-Methylnaphthalene	10.52	142	141, 115	0.999	2	-
1,6-Dimethylnaphthalene	11.97	156	141, 153	0.999	2	-
1,2-Dimethylnaphthalene	12.35	141	156, 115	0.999	2	-
Acenaphthylene	12.34	152	151, 150	0.999	2	-
Acenaphthene	12.75	153	154, 152	0.999	2	-
Fluorene	13.96	166	165, 167	0.999	2	65,300
Dibenzothiophene	15.91	184	139, 185	0.999	2	-
Phenanthrene	16.21	178	176, 179	0.999	10	490,000
Anthracene	16.33	178	176, 179	0.996	10	-
2-Methylphenanthrene	17.42	192	191, 165	0.998	10	-
2-Methylanthracene	17.53	192	191, 165	0.992	10	-
1-Methylphenanthrene	17.67	192	191, 165	0.999	10	-
9-Methylanthracene	18.00	192	191, 165	0.999	10	-
3,6-Dimethylphenanthrene	18.43	206	191, 205	0.999	2	-
Fluoranthene	19.01	202	203, 200	0.999	2	65,300
2,3-Dimethylanthracene	19.09	206	191, 205	0.996	10	-
9,10-Dimethylanthracene	19.60	206	191, 205	0.998	2	-
Pyrene	19.53	202	200, 203	0.999	2	49,000
Retene	20.27	219	220, 234	0.999	2	-
1-Methylpyrene	20.86	216	215, 217	0.999	2	-
Benz(a)anthracene	22.37	228	226, 229	0.983	10	350
Chrysene	22.45	228	226, 229	0.994	2	35,000
6-Methylchrysene	23.49	242	241, 226	0.999	2	-
Benzo(b)fluoranthene	24.78	252	253, 250	0.994	2	350
Benzo(k)fluoranthene	24.85	252	253, 250	0.992	2	3,500
Benzo(e)pyrene	25.44	252	250, 253	0.999	2	-
Benzo(a)pyrene	25.57	252	253, 250	0.996	2	35
Indeno(1,2,3-c,d)pyrene D-12	28.78	288	284, 289	Internal standard	-	-
Indeno(1,2,3-c,d)pyrene	28.86	276	277, 274	0.997	2	350
Dibenz(a,h)anthracene	28.95	278	279, 276	0.998	10	35
Benzo(ghi)perylene	29.63	276	277, 274	0.999	2	-
Dibenzo(a,i)pyrene	33.91	302	300, 303	0.998	2	-

Table 1. Summary of PAH GC-MS instrumental details and comparison to FDA levels of concern for finfish. *MDL = IDL x 2, where IDL was assigned when PAH lowest abundance SIM confirmation ion S/N > 3 for standards prepared in iso-octane.

Modified QuEChERS Experimental Design

Extraction Method	Solvent volume (mL) and composition	Salt amount (g) and type
Traditional E1 ^{b,c}	5; 1% acetic acid/acetonitrile	2.5; AOAC ^f
Traditional E2 ^{d,e}	2; acetonitrile	1.3; EN ^{g,h}
Modified E3	2; 2:2:1 ethyl acetate, acetone, iso-octane	1.3; AOAC
Modified E4	2; 2:2:1 ethyl acetate, acetone, iso-octane	1.3; EN

Table 2. Four QuEChERS methods compared on the basis of PAH extraction efficiency from smoked salmon. ^f AOAC salts – $MgSO_4$ and $Na_2C_2O_4$; ^{g,h} EN salts – $MgSO_4$, $NaCl$, $Na_2C_2O_4$, and $Na_2C_6H_6O_8$. Extraction methods E1 and E2 are based on published methods^{b,c,d,e}.

Modified -vs- Traditional QuEChERS Recovery Performance

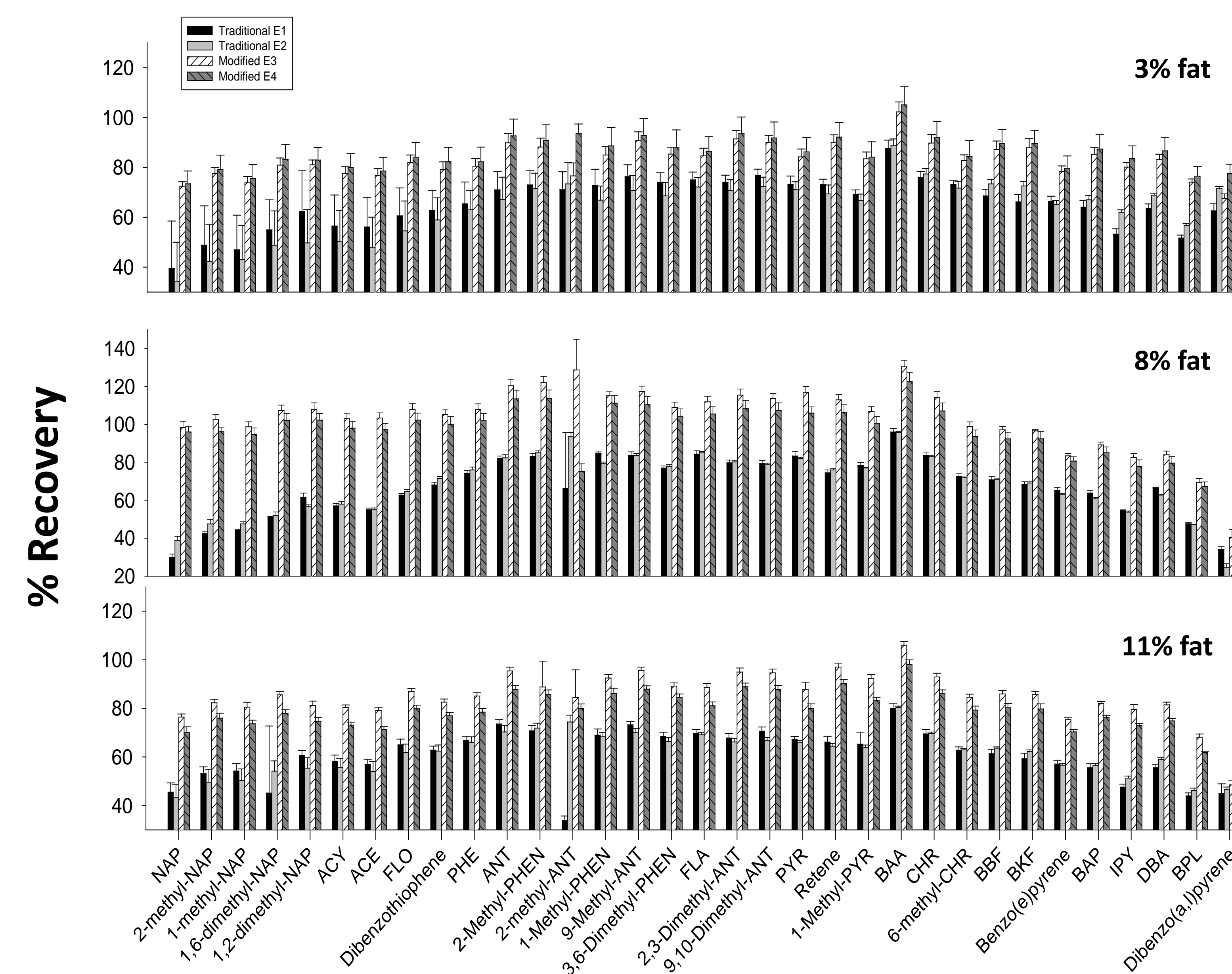


Figure 1. Average recovery of PAHs from smoked salmon (3 – 11% fat) generated with four different QuEChERS extraction methods (see Table 2 for details). Bars represent mean recoveries from four replicates ± SD.

Modified QuEChERS -vs- Soxhlet and ASE

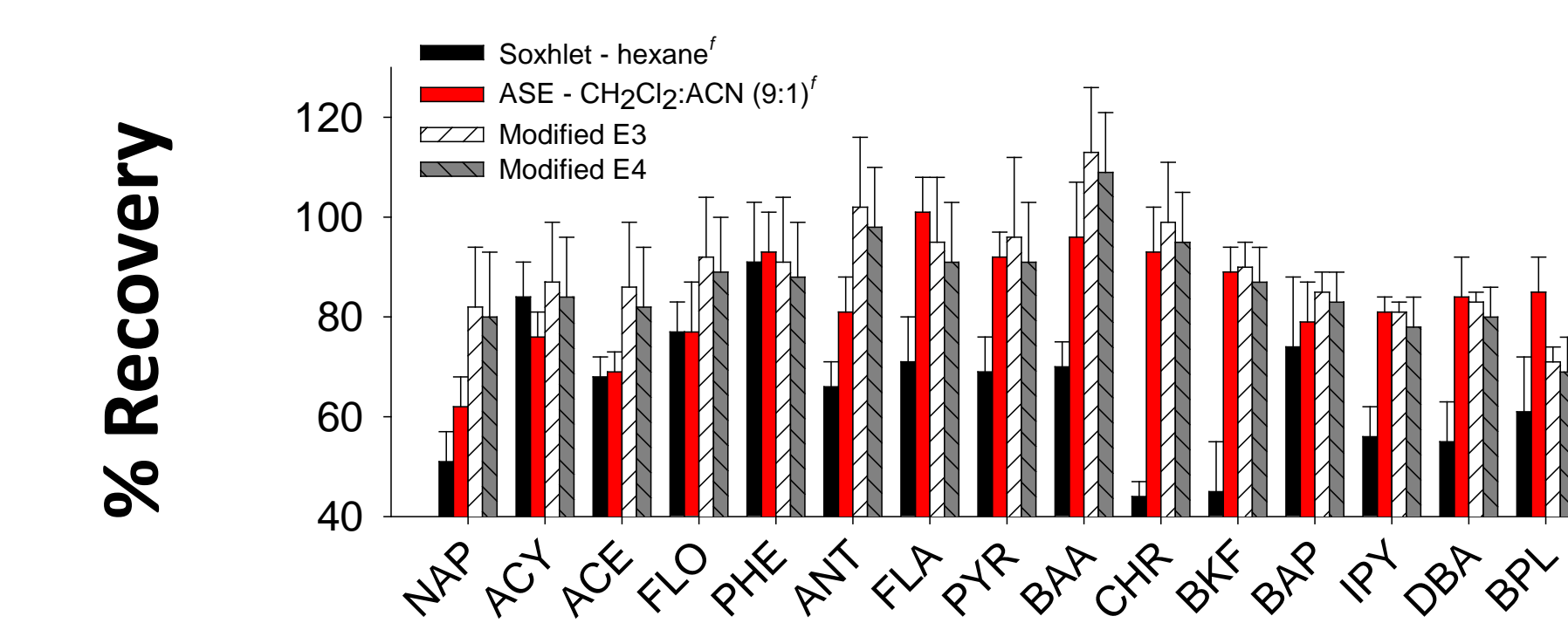


Figure 2. Comparison of PAH extraction performance (% recovery ± SD) for Soxhlet, accelerated solvent extraction and QuEChERS methods (E3 and E4). Soxhlet and ASE data are from Wang, 1999. E3 and E4 data represent mean PAH recovery across all three fish tested (n = 12/method).

Determination of PAHs in smoked salmon by Modified QuEChERS

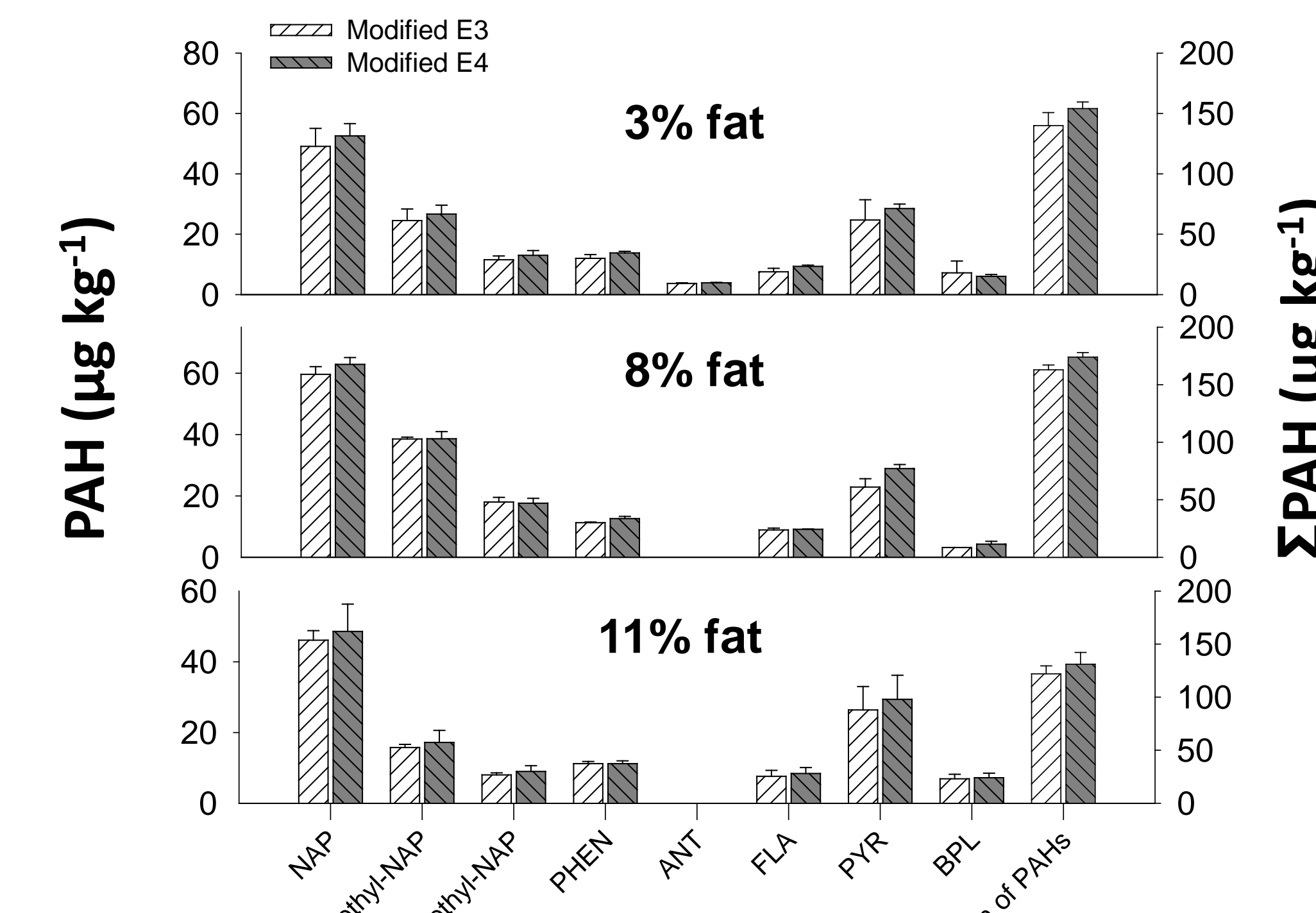


Figure 3. Comparison of PAH levels (mean µg kg⁻¹ wet weight ± SD, n = 3) measured in commercially available smoked salmon by two new QuEChERS extraction schemes (E3 and E4; see table 2 for details).

Conclusions

- A modified QuEChERS analytical method has been developed and validated for PAHs in high fat bio-matrices (Table 1 & Fig 1 – 3).
- Greater recoveries of PAHs are achieved with 2:2:1 ethyl acetate, acetone and iso-octane than acetonitrile alone (Figure 2).
- The type of partitioning salt used does not impact recoveries (Figure 1 and 3).
- QuEChERS methods perform comparably to, or better than, Soxhlet and Accelerated Solvent Extraction (ASE) based methods (Figure 2).
- Method sensitivity is sufficient for regulatory agencies (Table 1).
- Future investigations will apply the method to assess the contribution of traditional salmon smoking to dietary PAH intake for a Native American tribe.

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